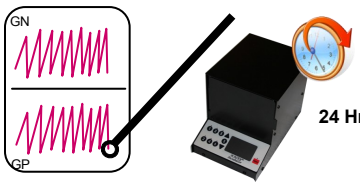
Kacey Diagnostics Quick Reference For Culture & Sensitivity Study

1 INNOCULATION OF CULTURE BI-PLATE

1) Remove the plate from refrigerator & pre-heat in the incubator @37 degrees for 15-20 minutes prior to inoculating the Kacey MultiChrome bi-plate.

2) Using a sterile inoculating loop (20uL) for liquids and a Kacey Sterile Swab for solids ,inoculate the sample onto both sides of the Kacey Multichrome Bi-plate, by utilizing a zigzag streaking motion.

3) Replace clear lid and place Kacey MultiChrome Bi-plate in the incubator **upside down** (inverted position) and incubate at 37°C +/- 2 degree C for no less then 24 hours . The plate should be examined after 24 hours, but no later than 48 hours after incubation.

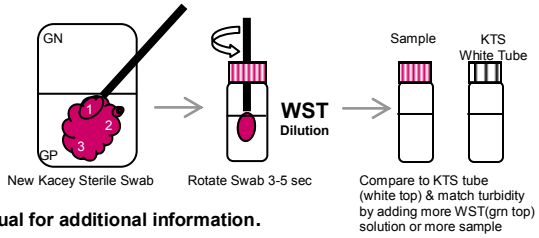


2 TRANSFER TO MULLER HINTON PLATE USING KACEY "WST" TUBES

1) Taking a Kacey Sterile Swab or Loop carefully dab into the three (3) different places containing the bacteria on the MultiChrome Bi-plate.

2) Immediately place the sterile Kacey Swab or Loop containing the bacteria into the Kacey Working Solution tube (**WST** which contains 1.0 ml of sterile 0.085% saline solution). Mix the Kacey Swab or Loop with a gentle twirling motion while in the **WST** tube for approximately 3-5 seconds.

3) Compare sample to the **Kacey Turbidity Standard** (white cap) **KTS**. Sample should match in turbidity, if not add more WST sol (green top) or more sample to match KTS tube. (see enclosed Turbidity Card).



3 INOCULATION OF MULLER HINTON (MH) PLATE

1) Using the Kacey Swab or loop containing the now diluted WST & Turbidity adjusted dilution streak the **1st time** the entire periphery of the Mueller Hinton plate making a 360° circle.

2) Streak a **2nd time** again the Muller Hinton Plate with a fresh sample of the WST dilution using wide broad strokes starting at the 9 to 3 o'clock position.

3) Streak a **3rd time** again the Muller Hinton plate with a fresh sample of the WST dilution using wide broad strokes starting at the 11 to 5 o'clock position

4) Remove "Sensi-Ring" from the foil pouch with tweezers at the inner tab & place the "Sensi-Ring" **facing down** onto the MH plate tapping down in non-disk areas, label the specimen to be incubated.

5) Place the MH Inoculated plate into an incubator **upside down,** set timer & incubate for 24 hours. Remove & read inhibition zones with Kacey Clear Acetate Reader.

Quick reference only. Refer to detailed instruction manual for additional information.

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Kacey Diagnostics
MICROBIAL ZONE INHIBTION INTERPRETIVE CHART*

w/ Zone Diameter Interpretive Standards
For Gram Positive, Gram Negative, Ear,
UTI and Skin/Wound Sensi-Rings™

Antimicrobial Agent Name	Sensi Code	Disc Potency	Resistant	Intermediate	Susceptible
Amikacin	AK30	30ug	≤14	7-9	≥ 17
Amoxicillin	A30	30ug	≤13	14-17	≥ 18
Augmentin-(Clavamox)	AUG30	30ug	≤13	14-17	≥ 18
Azithromycin-(Zithromax)	ATH12	15ug	≤13	14-17	≥ 18
- Staphylococcus spp.					
Ceftazadime	CAZ30	30ug	≤14	15-17	≥ 18
Cefpodoxime	CPD10	10ug	≤17	18-20	≥ 21
Cephalexin-(Keflex)	CFX30	30ug	≤14	15-17	≥ 18
Ciprofloxacin –(Cipro)	CIP5	5ug	≤15	16-20	≥ 18
Clindamycin –(Clincaps)	CD2				
- Staphylococcus ssp		2ug	≤14	15-20	≥ 21
- S.pneumoniae and other streptococci		2ug	≤15	16-18	≥ 19
Cotrimoxazole (trimethoprim/sulfa)	TS25	30ug	≤10	11-15	≥ 16
Choloramphenicol-(Chloromycetin)	C30	30ug	≤17	18-20	≥ 21
- Streptococci, staphylococci, enterococci, s.pneumoniae					
Cefalothin-(Keflin)	KF30	30ug	≤14	15-17	≥ 18
Doxycycline-(Vibramycin)	DXT30	30ug	≤12	13-15	≥ 16
Enrofloxacin-(Baytril)	ENF5	5ug	≤17	18-20	≥ 21
Gentamycin-(Gentocin)	GM10	10ug	≤12	13-14	≥ 15
Neomycin	NE30	30ug	≤12	13-16	≥ 17
Oxaciillin	OX10	10ug	≤10	11-12	≥ 13
Penicillin	PG10	10ug	≤28		≥ 29
Polymyxin	PB300	300ug	≤8	9-11	≥ 12
Ticarcillin-(Ticar)	TC75	75ug	≤14	15-19	≥ 20
Tobramycin-(Tobrex)	TN10	10ug	≤12	13-14	≥ 15
Trimethoprim/Sulfa-(Bactrim Septra Primor)	T/S25	25ug			
- Enterobacteriaceae, P.aeruginosa,					
- Acinetobacter,staphylococci			≤10	11-15	≥ 16
- s.pneumoniae					

*The Zone Inhibition Interpretive Chart values are the most widely accepted standard procedures that are used by the US Food and Drug administration (FDA) and the World Health Organization (WHO). These standard procedure values listed in the chart were adopted as a consensus standard by the Clinical and Laboratory Standard Institute (CLSI) formerly known as NCCL and is periodically updated. The latest CLSI documentation should always be consulted for current recommendations. The list of abbreviations for the antibiotics has been selected as standardization by KACEY® for worldwide sales, distribution, nomenclature and identification of the Kacey Sensi-Rings. **It is highly recommended that any bacterial agent whose etiology and treatment is beyond the scope of this system be sent to a qualified veterinary reference lab for more detailed testing.**

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